



# Surface phase transitions in foams and emulsions

Nikolai Denkov, Slavka Tcholakova and Diana Cholakova

## Abstract

Surface phase transitions in surfactant adsorption layers are known to affect the dynamic properties of foams and to induce surface nucleation in freezing emulsion drops. Recently, these transitions were found to play a role in several other phenomena, opening new opportunities for controlling foam and emulsion properties. This review presents a brief outlook of the emerging opportunities in this area. Three topics are emphasized: (1) the use of surfactant mixtures for inducing phase transitions on bubble surfaces in foams; (2) the peculiar properties of natural surfactants saponins, which form extremely viscoelastic surface layers; and (3) the main phenomena in emulsions, for which the surface phase transitions are important. The overall conclusion from the reviewed literature is that surface phase transitions could be used as a powerful tool to control many foam and emulsion properties, but we need deeper understanding of the underlying phenomena to fully explore these opportunities.

## Addresses

Department of Chemical and Pharmaceutical Engineering, Faculty of Chemistry and Pharmacy, Sofia University, 1 James Bourchier Avenue, Sofia, 1164, Bulgaria

Corresponding author: Denkov, Nikolai ([nd@lcpe.uni-sofia.bg](mailto:nd@lcpe.uni-sofia.bg))

Current Opinion in Colloid & Interface Science 2019, 44:32–47

This review comes from a themed issue on **Memorial Volume**

Edited by **Debora Berti, Dganit Danino, Peter A. Kralchevsky, Rene Nome, Elena Mileva and Larry Romsted**

For a complete overview see the [Issue](#) and the [Editorial](#)

<https://doi.org/10.1016/j.cocis.2019.09.005>

1359-0294/© 2019 Elsevier Ltd. All rights reserved.

## Keywords

Surface phase transition, Liquid condensed phase, Liquid expanded phase, Adsorption layer, Foam, Emulsion, Partial coalescence, Surface nucleation, Drop self-shaping, Saponin.

## Introduction: role of surface ‘rigidity’ in foam studies

The role of surface ‘rigidity’ in the mode of foam film thinning was first described by Miles et al. [1] and then elaborated in great detail in the classical book ‘Soap Films’ by Mysels et al. [2]. In a series of experimental observations with solutions of different surfactants, these authors revealed several modes of foam film thinning, and the terms ‘rigid’ and ‘mobile’

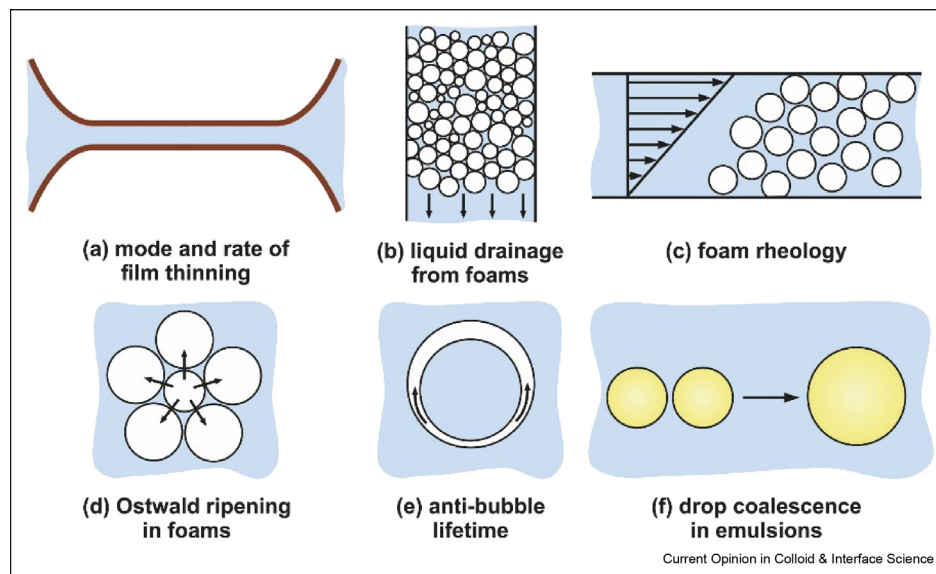
films were introduced for the two extreme types of behavior. Epstein et al. [3] pointed out that surface rigidity is probably due to formation of ‘solid mixed surface layers’ and emphasized the role of the dodecyl alcohol ( $C_{12}OH$ ) as an essential admixture that rigidifies the adsorption layers of sodium dodecyl sulfate (SDS), which was used as a main surfactant in most experiments. Thus, the intricate link between the phase state of surfactant adsorption layers and the film-thinning behavior was recognized for the first time.

In the following years, several groups developed theoretical approaches to quantify the relations between the viscoelastic properties of the adsorption layers and foam film-thinning behavior. Ivanov et al [4] analyzed in detail the role of surfactant mass transfer and the related Marangoni effect, which creates apparent surface elasticity. Langevin [5] analyzed the role of surface viscosity. Joye et al. [6] explained the role of surface viscoelasticity in the phenomenon of marginal regeneration observed in thinning foam films. Somewhat surprisingly, neither of these studies tried to establish a deeper connection between the film-thinning processes and the wide research area of surface phase transitions in spread surfactant layers, which was well established in the literature by that time (see [Surface phase transitions at the air–water interface in relation to foam properties](#)).

It took almost 40 years after the book ‘Soap Films’ [2] was published, when it became clear that the surface phase transitions have a profound impact on many other dynamic properties of foams (see [Figure 1](#)). First, in the studies of Bergeron on the coalescence stability of foam films, he [7] showed that surface dilatational elasticity might be an equally important factor for film stabilization, along with the well-known surface forces (electrostatic, van der Waals, and so on).

Second, an inspiring debate about the rate of liquid drainage from foams [8,9] ended with the conclusion that two different regimes are possible, depending on the surface mobility of foam bubbles. By adding lauryl alcohol to SDS solutions, Stoyanov et al. [10] changed the regime of liquid drainage, thus demonstrating the direct link to the surface phase transitions. Theoretical models were developed to describe the regimes of ‘rigid’ and ‘fully mobile’ interfaces [11,12].

Figure 1



Schematic presentation of several phenomena for which the surface phase transitions in the surfactant adsorption layers change the foam or emulsion behavior very significantly: **(a)** Mode and rate of film thinning. **(b)** Liquid drainage from foams. **(c)** Foam rheology. **(d)** Ostwald ripening in foams. **(e)** Lifetime of anti-bubbles. **(f)** Drop coalescence in emulsions.

Third, in a series of articles, Denkov *et al.* [13,14] showed that the foam rheological response also exhibits two qualitatively different regimes, controlled by the surface mobility of the bubbles. Notably, it was shown for the first time in these studies [13–15] that the oscillating drop method is a very convenient tool to characterize the surface mobility. High surface moduli  $E > 100$  mN/m, as measured at low oscillation frequency (0.1–1 Hz), always corresponded to the regime of ‘rigid’ interfaces, whereas  $E < 50$  mN/m always corresponded to mobile interfaces [15]. The long-chain fatty acids (C14 to C18) were introduced in these studies as convenient cosurfactants to induce surface phase transition (condensation).

In a parallel series of experiments, Stocco *et al.* [16], Varade *et al.* [17], and Tcholakova *et al.* [18] studied the role of surface elasticity on the bubble Ostwald ripening in foams. Both series of experiments concluded that bubble ripening is much slower at high surface elasticity. Two complementary explanations were given to this effect [16–18]. Both the surface elasticity and reduced gas permeability of the condensed adsorption layers could play a role in the strong reduction of the Ostwald ripening rate, and the relative importance of these two effects is system specific.

The various effects mentioned previously have been studied multiple times in the following years by various research groups, which confirmed the important role of surface condensation in all dynamic properties of foams. Thus, the role of surface rigidity/mobility in foams has

been firmly established. Nevertheless, a systematic study about the relation between the foam properties and the various possible surface phases in the adsorption layers (see [Main surface transitions and experimental methods for their investigation](#)) has not been conducted so far.

The role of surface phase transitions in the adsorption layers covering the oily drops was also studied by several groups in the context of drop crystallization and drop–drop partial coalescence in cooled oil-in-water emulsions [19–21].

The major aim of this review is to summarize the recent progress in the area of surface phase transitions in foams and emulsions. The important open questions and possible directions for future research are outlined.

Owing to the limited space, the focus of this review is exclusively on the adsorption layers of low-molecular-mass surfactants (including saponins). The adsorption layers of solid particles and polymer molecules (including the biopolymers, such as proteins and polysaccharides) exhibit a large variety of additional, strongly system-specific, effects that are not reviewed here.

### Surface phase transitions at the air–water interface in relation to foam properties

#### Main surface transitions and experimental methods for their investigation

It is accepted to call ‘Langmuir monolayers’ the spread surface monolayers of insoluble molecules on a liquid

subphase. In contrast, the monolayers formed via spontaneous adsorption of dissolved amphiphilic molecules from adjacent bulk phases are called ‘Gibbs adsorption layers’ [22]. For brevity, we also use this notation in the following section.

Historically, the surface phase transitions were first observed in Langmuir monolayers. Using the Langmuir trough to vary precisely the area per molecule in the layer and measuring the surface tension by the Wilhelmy plate method allowed the researchers to detect the main surface phase transitions and to clarify the role of the main factors involved. Typically, the surface phase transitions follow (part of) the sequence gas  $\rightarrow$  liquid expanded  $\rightarrow$  liquid condensed  $\rightarrow$  solid ( $G \rightarrow LE \rightarrow LC \rightarrow S$ ) as the temperature and/or the area per molecule is reduced (see Figure 2) [23–25]. The specific temperatures and adsorptions, at which these surface phase transitions are observed, depend strongly on the head group, chain length, branching, and presence of double bonds in the surfactant tail. A summary of these classical studies can be found, for example, in the books by Gaines [23] and Adamson and Gast [24].

More recently, many powerful structural methods were developed or adapted for surface studies, and they revealed a rich variety of polymorphic structures formed at the air–water interface. The invention of monolayer-sensitive microscopy techniques, such as fluorescence microscopy, Brewster angle microscopy [26], and polarized fluorescence microscopy [27], revealed mesoscopic

structures of peculiar shapes in Langmuir monolayers. The fine molecular structure of the surface layers was characterized by synchrotron techniques, including X-ray scattering, X-ray reflectivity, grazing incidence X-ray diffraction, and neutron scattering and reflectivity, and by various spectroscopy techniques, such as surface-specific sum frequency generation spectroscopy and infrared reflection–absorption spectroscopy [28]. Atomic force microscopy of Langmuir–Blodgett layers transferred onto solid substrates was also used to analyze the structure of Langmuir monolayers and of more complex surface aggregates and multilayers [29].

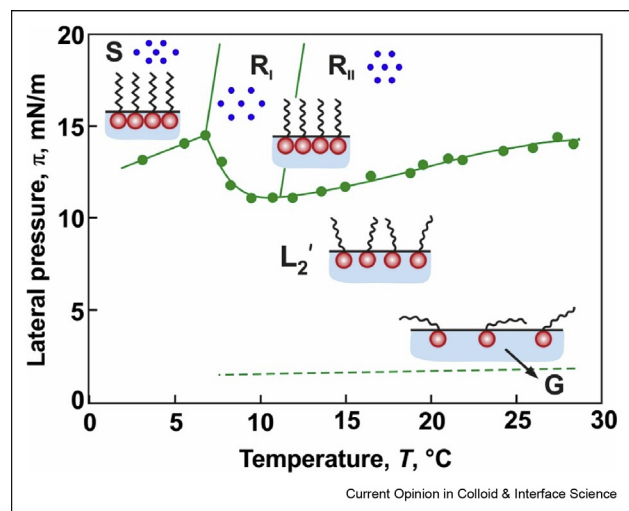
During the last years, surface rheological measurements using the oscillating drop method [30], capillary pressure tensiometer [31], magnetic needle [32,33], and double-wall ring method [34] provided valuable complementary information about the phase transitions in Langmuir and Gibbs layers. For example, a significant increase in the surface viscosity was observed on the phase transition  $LE \rightarrow LC$  in tricomponent DPPC:PA:Chol monolayers, studied at different surface pressures and cholesterol fractions [35\*\*]. Recent reviews on the methods for characterization of the surface rheological properties are presented in the studies by Langevin and Monroy [36] and Langevin [37].

It is worth mentioning that several recent studies [38–40] warned about possible problems with some of these experimental methods. Thus, it was shown that Langmuir trough–based techniques may give unreliable results for the reversibility of surface phase transitions, when reaching low surface tensions, owing to the leakage of material [38]. An approach to overcome this problem was proposed in the study by Xu and Zuo [39] where reversible phase transition in phospholipid layers was studied using a new method, called ‘constrained drop surfactometry,’ which is claimed to provide a leakage-proof environment.

In another study, Cuenca et al. [40] resolved the previous puzzling reports about the measurements of apparently negative surface viscosity in some surfactant systems. The authors found that the surfactant adsorption–desorption processes might be highly asymmetric, with different characteristic times and mechanisms. This asymmetry invalidates the accepted conventional procedure for data analysis that leads to physically unrealistic negative values of surface viscosity.

Because the Langmuir layers of insoluble surfactants are inefficient for stabilizing foams and emulsions, in the following section, we focus our review on the Gibbs layers, formed via spontaneous adsorption of surfactant molecules from the adjacent surfactant solution. Several approaches have been applied to induce phase transitions in Gibbs monolayers, as illustrated in Figure 3, and described in the following section.

Figure 2



Schematic presentation of the main surface phases, observed in octadecanol monolayers spread on water surface. G denotes the gas phase,  $L_2'$  denotes the liquid expanded (disordered) phase,  $R_I$  and  $R_{II}$  denote the two liquid condensed (rotator) phases, and S denotes the crystalline solid phase. The various phases differ in the area per molecule, tilt of the alkyl chain, and in-plane structure, as shown in the figure (adapted from the study by Lautz et al [25]).

### Mixtures of soluble and insoluble surfactants

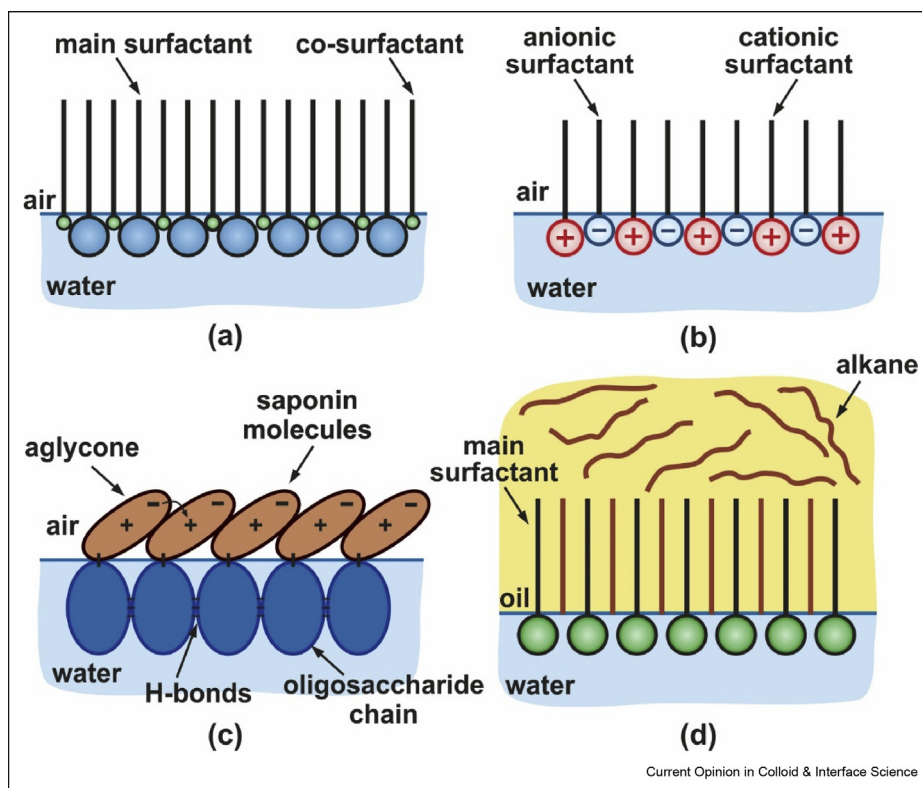
The solubilization of water-insoluble surfactants in the micelles of water-soluble surfactants has proven to be a powerful approach for forming a mixed adsorption layer that can undergo surface phase transitions with the formation of highly viscoelastic surfaces (see [Figures 3a and 4](#)). Usually fatty alcohols or fatty acids are used as cosurfactants to various anionic or cationic main surfactants [15–17,41–43\*]. Several recent studies are reviewed in the following section to illustrate the universal character, the versatile use, and the importance of this approach.

In the study by Wang *et al.* [42], it was shown that the addition of C<sub>12</sub>OH to SDS-containing foaming solutions increases significantly the foam stability owing to the increased viscoelasticity of the formed mixed adsorption layers. In contrast, the foam stability is governed mostly by electrostatic interactions (and is lower) at the same concentration of SDS, in the absence of C<sub>12</sub>OH. These

results are another convincing support to the concept [7] that surface viscoelasticity plays a very important role in foam stabilization.

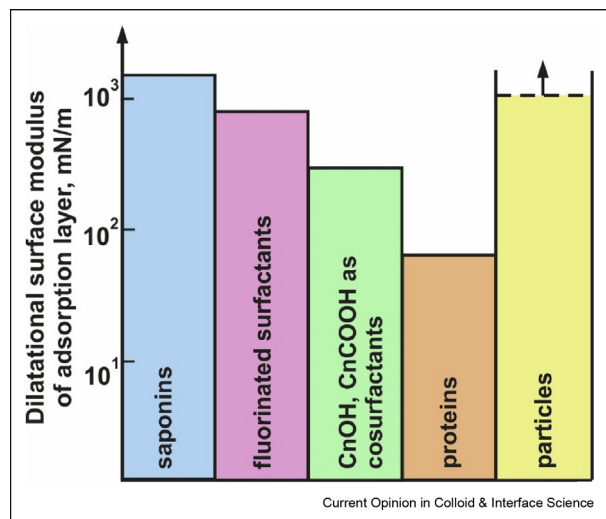
In a recent study with mixed solutions of myristic acid (MAc) and choline hydroxide [44\*], it was shown that the pH and ratio of the two components play a key role in foam stabilization. The choline is used as a bulky counterion that increases the solubility of MAc at low pH. The authors found that foam stability is maximum at pH values around the pK of MAc. Under these conditions, dense mixed adsorption layers of ionized and nonionized species of MAc are formed at the bubble surface and stabilize the foam very efficiently. In other words, the adsorption layer behaves as composed of two different surfactant components, one of them being a fatty acid with small head group, while the other is the ionic form of this acid. Thus, the layer structure closely resembles the one shown in [Figure 3a](#), despite the fact that only one surfactant substance is dissolved initially.

**Figure 3**



Schematic illustration of the main approaches used to induce surface phase transition in adsorption layers of low-molecular-mass surfactants at air–water and oil–water interfaces. **(a)** Addition of poorly water-soluble cosurfactants with small head groups. The cosurfactants are solubilized in the surfactant micelles and form mixed adsorption layers with the main surfactants at the air–water interface. **(b)** Mixing anionic and cationic surfactants in an appropriate ratio so that the surface complex of the two surfactants is formed. Alternatively, multivalent counterions could be used to induce condensation of the adsorption layer of charged surfactant molecules and even to form surface multilayers of these molecules. **(c)** Saponin molecules pack well at air–water and oil–water interfaces owing to strong attraction between their large hydrophobic aglycons and owing to multiple hydrogen bonds between their hydrophilic sugar groups. **(d)** Interdigitating long-chain alkane molecules into adsorption layers of long-chain surfactants leads to formation of mixed alkane–surfactant monolayers that may freeze at temperatures higher than the freezing temperature of the adjacent bulk phases.

Figure 4



Typical range of the dilatational surface moduli of adsorption layers of water-soluble low-molecular-mass surfactants as measured at low oscillation frequencies (ca. 0.1–1 Hz) in comparison with adsorption layers of common proteins and with particle interfacial layers.

In a systematic series of experiments, the effects of chain length and concentration of fatty acids (used as cosurfactants), pH, and temperature on the onset of surface phase transition and on the related foam behavior were studied by Mitrinova et al. [15,43\*]. Several important conclusions were drawn: (1) Most viscoelastic layers at 21 °C were obtained from myristic and palmitic acids having chain lengths of C14 and C16, respectively. The longer stearic acid with 18 carbon atoms was not solubilized well in the micelles of the main surfactants, and hence, the viscoelasticity of the respective mixed adsorption layers was lower than that of the myristic and palmitic acids. (2) Increasing temperature and/or pH can be used to melt the surface layers, which leads to jump-wise decrease in the surface viscoelasticity by ca. 2 orders of magnitude. (3) Neutron reflectivity measurements showed that (a) the cosurfactant molar fraction in the adsorption layer is comparable with that of the main surfactant, despite the much lower cosurfactant concentration in the bulk solution, and (b) the viscoelastic layers correspond to an average area per molecule of  $\approx 0.20 \text{ nm}^2$  (unpublished results). This latter value evidences that the molecules in the viscoelastic layers have the area per molecule resembling that in the intermediate rotator phases, observed with bulk alkanes, rather than the area corresponding to the fully crystalline structure of alkyl chains which is  $< 0.19 \text{ nm}^2$  [45]. (4) Well-defined boundary exists in the surface viscoelastic modulus, between 50 and 100 mN/m (depending on the specific foam phenomenon studied), which leads to a qualitative shift in the

foam behavior and thus separates the regimes of rigid and fluid interfaces.

Marinova et al. [46] applied this approach to even more complex mixtures of surfactants, anionic sodium lauryl ether sulfate (SLES) + nonionic alkyl polyglycoside (APG) + zwitterionic cocamidopropyl betaine (CAPB). The addition of C<sub>12</sub>OH was found to decelerate the rate of water drainage from the respective foams at high salt concentration and low temperature. These results were explained by the higher viscoelasticity of the C<sub>12</sub>OH-containing adsorption layers.

Several articles reported that surface viscoelasticity plays a role in foam systems relevant to enhanced oil recovery. Thus, Wang et al. [47] showed that the addition of long-chain fatty acids, which increase the surface viscoelasticity of adsorption layers, could strongly enhance the foam tolerance to the antifoam action of oils. In an independent study, Mensire et al. [48] showed that the rheological properties of the interfaces and of the respective foams strongly affect the oil extraction parameter from porous rocks. Along the same line of reasoning, Wang et al. [49,50] showed that higher surface dilatational modulus decreases the surface deformability and, thus, impacts the foam flow in the porous medium.

Several studies showed that the surface viscoelasticity is very important for other foam-related phenomena as well. Dollet and Bocher [51] showed that the surface viscoelasticity changes qualitatively the dynamic behavior of a bubble monolayer (2D foam) flowing between two solid plates. These results were explained by considering the role of surface viscoelasticity on the bubble-wall friction, bubble elastic deformation, and the rate of bubble plastic rearrangement. Vitry et al. [52] showed that surface elasticity is a key factor to increase the lifetime of the so-called ‘antibubbles’ — spherical gas films, freely floating in surfactant solutions (Figure 1e).

#### Mixtures of oppositely charged surfactants (catanionic surfactants) and of ionic surfactants with multivalent counterions

In a series of articles, Langevin et al [16,17] studied the foaming properties of mixtures of cetyl trimethylammonium bromide (CTAB) and MAc, considered to be composed of oppositely charged surfactants. In the studies by Stocco et al [16] and Varade et al [17], they showed that mixed adsorption layers with low surface tension and high surface elasticity can be formed. The respective foams were very stable to bubble Ostwald ripening and bubble–bubble coalescence. The surface properties of these solutions were characterized in detail in the study by Arriaga et al [53\*]. The existence of a melting temperature was reported above which the

surface layer was fluidized, similar to the observations in the studies by Mitrinova et al [15,43\*].

As summarized in the very informative review by the same research group [54\*], the formation of such highly viscoelastic surface layers, using mixtures of oppositely charged surfactants, is often combined with other mechanisms leading to ultrastable foams, such as gelation of foam films and/or bulk foaming solution. A recent example of such an approach was presented in the study by Zhang et al [55], where precipitation of the surfactant on the bubble surface was used to form particle-stabilized (Pickering) ultrastable foams, responsive to temperature variations.

Lyadinskaya et al. [56\*\*] showed that the surface tension of CTAB, dodecyl trimethylammonium bromide (DTAB), and DNA solutions does not change, if these solutes are dissolved separately at very low concentrations. However, if DNA and cationic surfactants are dissolved together at the same concentrations, strong synergistic effect is observed, and the surface tension decreases owing to the formation of DNA–surfactant complexes. The surface elasticity of the mixed DNA–CTAB solution increases up to 85 mN/m, while the maximal value for CTAB alone is  $\approx 20$  mN/m. The authors explained their results with combined strong electrostatic and hydrophobic attraction between the DNA and surfactant molecules, which leads to surface complexation. The hydrophobic attraction is between the hydrophobic surfactant tail and the DNA nucleobases exposed in the major groove of the double helix. Several other interesting phenomena were described in these systems, such as transition from a loose network to dense and brittle adsorption layer, formation of adsorption multilayers, and growth of fibrillar aggregates, which can merge and form continuous surface phase [56\*\*].

Another approach to induce surface phase transitions using appropriate control of the electrostatic interactions is to add multivalent counterions to ionic surfactants, which can lead to surface ordering and even formation of surfactant multilayers at the interface. Such multilayers were detected recently by neutron reflectivity on addition of multivalent cations to solutions of the anionic surfactants  $\alpha$ -methyl ester sulfonate [57–59] and SLES [60].

### Saponin adsorption layers and saponin-stabilized foams

Saponin is a generic term for a large class of natural amphiphilic molecules that are widely spread in the plant

kingdom and can be extracted at the industrial scale from many plant species. Some of the saponin extracts form highly viscoelastic adsorption layers at the air–water interface and could be very efficient foam stabilizers. Therefore, studies on the saponin surface and foaming properties appear regularly during the last years.

The high viscoelasticity of the saponin adsorption layers is caused by strong attraction between the adsorbed saponin molecules that leads to surface phase transition, as shown by analysis of surface tension isotherms of saponin solutions [61\*]. Molecular dynamic simulations [62\*] showed that the intermolecular attraction includes strong dipole–dipole interactions and hydrogen bonding. Some saponins form elastic ‘skin’ at the air–water and oil–water interfaces with strongly nonlinear response on surface deformation [63,64].

It is worth noting that the commercial saponin extracts are usually multicomponent mixtures, containing molecules with different solubility in water [61\*,65,66]. Therefore, one expects that the formation of viscoelastic adsorption layers for many of these extracts could be affected by the presence of both highly water-soluble components (which form micelles) and poorly water-soluble components (which could be solubilized inside the micelles). These different components could form mixed adsorption layers with high viscoelasticity, just as the more conventional surfactant mixtures discussed in the [Mixtures of soluble and insoluble surfactants](#) section.

Wojciechowski [65] and Golemanov et al. [66] clarified that surface properties of saponin solutions may differ significantly, depending on their source and producer. Kezwon and Wojciechowski [67\*] also studied mixed adsorption layers of *Quillaja* saponin (QS) with proteins and lipids, and strong surface complexation was reported for some of these systems. In a separate study, Böttcher et al. [68] showed that QS and the globular milk protein beta-lactoglobulin also form complexes at the air–water interface with strong synergistic effect on foam stability.

Ulaganathan et al. [69] studied the surface and foam film properties of solutions of QS. The effect of pH was found to be relatively small for the surface properties and for the rate of film thinning. The final film thickness, however, was affected by pH — an effect attributed to changed electrostatic repulsion between the foam film surfaces.

Pagureva et al. [61\*] found a clear relation between the surface viscoelasticity of the saponin adsorption layers and the rate of bubble Ostwald ripening in the respective foams. Two properties of the adsorption layers were

found to be important for the decelerated bubble ripening in saponin-stabilized foams. First, the condensed saponin adsorption layers have very low permeability for the gas molecules, which should cross the foam films in the process of gas exchange between the bubbles (Figure 1d). Second, the surface tension of the shrinking bubbles could be significantly lower than the equilibrium one, thus decreasing the bubble capillary pressure that is the main driving force for gas transfer. These two combined effects reduce the rate of bubble Ostwald ripening by up to 2 orders of magnitude compared with the foams stabilized by conventional surfactants.

The foamability and foam density for solutions of different saponins were compared in the studies by Böttcher and Drusch [70,71]. However, these authors could not find clear correlations between foam properties and the measured surface characteristics.

Enhanced foam stability after addition of chitosan to saponin solutions was reported in the study by Santini et al [72], especially with respect to the suppressed bubble coalescence during foaming. This effect was partially explained by the increased viscosity of the mixed saponin–chitosan solutions, which leads to the formation of smaller and more stable bubbles.

Two new extracts of saponins from the plant species *Quillaja brasiliensis* and *Agave angustifolia* were studied by Müller and Schiedeck [73]. The authors found that the surface and foaming properties of the studied saponin solutions depended strongly on the method of extract preparation, which is in agreement with previous studies.

Stable foams were formed by solutions of glycyrrhizic acid (GA) in which saponin nanofibrils were formed and stabilized the bubbles [74].

One sees from the aforementioned brief review of the recent publications that the saponins are of high research interest. However, the complexity of their surface properties and the variability of the studied saponin extracts make the physicochemical interpretation of the obtained results very difficult. One could expect a slow but steady progress in this area.

### Fluorinated surfactants

Several recent studies showed that the fluorinated surfactants could be very efficient in forming highly viscoelastic adsorption layers and in enhancing foam stability. Thus, Kovalenko et al. [75\*\*] showed that fluorinated surfactants, with phosphonic head groups and 8–10 carbon atoms in their chains, form adsorption layers with very high elasticity, up to 950 mN/m. High elasticities, up to 250 mN/m, were also reported in the

study by Noskov et al [76\*] for adsorption layers of heptadecafluoro-1-nonanol ( $C_9H_{17}F_{17}OH$ ). Phase transition of the type LE  $\rightarrow$  LC in the adsorption layer was reported, and the high surface viscoelasticity was attributed to the LC phase. An independent study of the short-chain zwitterionic fluorocarbon cosurfactant FS50, added to SDS solutions, showed very strong synergistic effects and enhanced foam stability for the mixed solutions [77].

Obviously, the foaming properties of the fluorinated surfactants and their mixtures with the common alkyl-based surfactants are poorly understood as we do not know the detailed mechanisms governing these surface phase transitions and leading to the reported very high surface viscoelasticities. New interesting results could be expected in this emerging research area.

### Stimulus-responsive surfactants and smart foams

There are rapidly growing interest and research activity in stimulus-responsive foams and emulsions. In a number of studies, temperature and pH were successfully used to modify the surface and foaming properties of various surfactant solutions in a desired way [78–82].

A new approach for foam control was developed recently, which requires specially synthesized photosensitive surfactants with cis–trans isomerization of the surfactant molecules and relies on the surface phase transitions triggered by light illumination. Chevallier et al. [79–81\*\*] characterized the surface and foam film properties for solutions of such light-sensitive surfactants. Several unexpected phenomena were observed owing to nontrivial Marangoni effects and to changes in the surfactant aggregates when the surfactant conformations were shifted by light illumination [79–81\*\*].

Independently, Jiang et al. [82] showed that the foaming ability of the solutions of the 4-butyl-4'-(4-N,N-dimethylbutoxyamine) azobenzene bicarbonate (AZO-B4) light-sensitive surfactant is comparable with that of  $C_{16}TAB$  and that the solution properties could be controlled using UV illumination and/or pH variation of the solution, for example, via purging of  $CO_2$  through the solution.

Certainly, this area will be expanded in the following years because the stimulus-responsive systems will remain of high interest from both academic and applied viewpoints.

### Phase transitions at the oil–water interface and their effect on emulsion properties

One could expect that, similar to foams, the phase transitions in the surfactant adsorption layers, formed at oil–water interfaces, should affect the emulsion

properties significantly. Until recently, however, these effects have not been systematically studied, with one exception — in relation to the partial coalescence of the oily drops in cooled oil-in-water emulsions, when the drops undergo transition from the liquid phase to solid phase.

Arguably, the main reason for this limited interest in surface phase transitions in emulsions is that the central approach to induce surface phase transitions in foams (namely, via addition of water-insoluble cosurfactants with small head groups, Figure 2a) is not applicable to emulsions. Indeed, the long-chain fatty alcohols and acids are highly soluble in the oil phase. Therefore, no condensed

(frozen) adsorption layers are formed in emulsions under conditions that would produce such layers in foams.

Two other main approaches were found useful to induce surface phase transition at the oil–water interfaces with possible application to emulsions, as briefly explained in the following section.

#### Long-chain trimethylammonium bromide and chloride surfactants

The first approach relies on using long-chain trimethylammonium bromides or chlorides ( $C_n$ TAB or  $C_n$ TAC with  $n = 14–18$ ) [83–86\*]. The relatively large charged head group of these surfactants suppresses their solubility in the oil phase. Thus,  $C_n$ TAB or  $C_n$ TAC has very high affinity to the oil–water interface and readily forms relatively dense adsorption layers. Interfacial freezing of these layers was observed only when alkanes with chain length comparable with that of the surfactant tail were present in the hydrocarbon phase (as single constituents or in alkane mixtures). The experiments evidenced that the frozen adsorption layers contain surfactant and interdigitated alkane molecules that nicely pack together on layer freezing [86\*] (see Figure 3d).

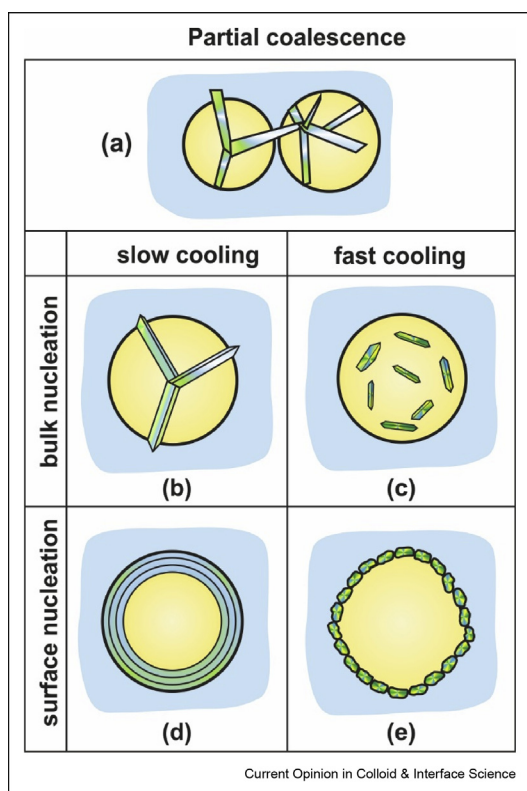
The series of articles by Wilkinson [84], Sloutskin *et al.* [85], Yefet *et al.* [86\*], Matsubara *et al.* [87], and Tokiwa *et al.* [88] revealed the role of various factors in the interfacial phase transitions at the oil–water interface and opened the door for the study of their effect on emulsion properties. A very comprehensive review on the progress in this area was published recently by Matsubara and Aratono [89\*\*].

Following this line of research, Tokiwa *et al.* [90\*] demonstrated that alkane-in-water emulsions, stabilized by cetyl trimethylammonium chloride surfactant (CTAC), remained stable for a period of 24 h, when stored at temperatures corresponding to the frozen adsorption layer on drop surfaces. In contrast, at temperatures higher than the layer melting temperature, the emulsion became unstable — the bulk oil phase separated immediately after emulsification, and the emulsion volume decreased continuously with time owing to drop–drop and drop–homophase coalescence. The authors explained this trend with the higher mechanical elasticity of the frozen adsorption layers and the related slower drainage of the emulsion films, formed between the neighboring droplets [90\*].

#### Long-chain monoglycerides and polysorbates in emulsions undergoing partial coalescence

The second approach was proposed in the studies of the partial coalescence phenomenon in cooled emulsions (Figure 5). It includes the use of long-chain monoglycerides or polysorbates (Tweens or Spans), which may induce interfacial freezing in such emulsions.

Figure 5



Schematic presentation of the effects of the crystal nucleation site (bulk or surface) and cooling rate (fast or slow) on the type of crystals formed and on the partial drop–drop coalescence in cooled emulsions. (a) Two droplets in close proximity: the growing lipid crystals may break the aqueous film between them, thus causing partial drop–drop coalescence. (b) Large crystals, protruding from the drop interior, are formed in the case of bulk nucleation and at a slow cooling rate. (c) Smaller crystals, which remain trapped inside the drop interior, are formed on rapid cooling in bulk nucleation. (d) Large crystals grow from the drop surface inward when slow cooling and surface nucleation are combined. (e) Crust of small crystallites is formed first in the case of surface nucleation and fast cooling. Owing to the subsequent oil shrinkage on freezing of the remaining oil in the drop interior, this crust can be distorted, and crystal bulges can be formed on the surface of the freezing drop. Cases (b) and (e) are proposed in the literature as promoting partial coalescence, whereas cases (c) and (d) are considered as less efficient in inducing partial drop coalescence [95,96].

The partial coalescence occurs in the temperature range in which the dispersed oily phase is partially crystallized [91]. The process is triggered by the solid crystals that appear in the droplets on emulsion cooling [92,93]. These crystals may protrude from the freezing droplets into the continuous phase and on collision with another droplet, which might be still in a liquid state owing to the supercooling phenomenon [91]; they may puncture the emulsion film separating the neighboring drops (Figure 5a). The created drop–drop contacts induce partial or complete freezing of the second drop because the protruding crystals act as nuclei for crystal growth within the volume of the second drop. The rapid freezing of the contacting drops precludes their complete merger, which justifies the used term ‘partial coalescence.’ When many drop–drop contacts are formed on storage or during shearing of the cooled emulsion, a 3D network of interconnected solid or semisolid particles is formed, and the disperse system increases its apparent viscosity very significantly, up to possible complete gelation [19,91,94].

Partial coalescence was studied in relation to several applications. In food systems, partial coalescence might be a desired phenomenon, if a stable semisolid dispersion with a desired plastic texture is to be produced (ice cream, whipped toppings, butter, and so on) [91,94]. In contrast, partial coalescence should be avoided during the long-term storage (shelf life) of sauces, creams, and other dairy products. Partial coalescence could also appear as a severe technological problem, if it occurs prematurely, as the rheological properties of the partially coalesced dispersion may become inappropriate for its downstream processing. Particle coalescence is also an important technological problem when formulating stable alkane-in-water emulsions for energy storage and transportation (emulsions of phase change materials), in which the latent heat of the transition from the liquid phase to solid phase of the alkane drops is used as an energy reservoir [19].

Therefore, the main factors of control of partial coalescence have been studied widely, including the process parameters during emulsion preparation and storage, oil volume fraction and chemical composition, chemical composition of the continuous phase, drop size, and composition of the interfacial layer [92–94]. The results reported in the various articles are often in apparent contradiction to each other. These contradictions are briefly described below and a plausible unifying explanation, based on the role of surface freezing, is proposed.

Fat crystals protruding out of the surface of fatty droplets, which are able to penetrate into the aqueous phase and to break the emulsion films, were visualized in some experiments with large drops (15–20  $\mu\text{m}$ ), stabilized by polysorbate 20 (Tween 20) with 12 carbon atoms in the surfactant chain [95]. However, Moens et al. [96] did

not confirm the occurrence of well-defined penetrating fat crystals in the partial coalescence phenomenon.

To explain the absence of such protruding crystals in their experiments, Moens et al. [96] proposed that partial coalescence may involve the occurrence of local heterogeneities (corrugations) in the surface crust of frozen fat, instead of being caused by distinct fat crystals, protruding from the drop surface. These authors hypothesized that the surface crystalline crust may deviate from the initial spherical shape of the drop surface owing to oil shrinkage on drop freezing, resulting in the formation of frozen fat bulges on the drop surface (Figure 5e), which may induce partial drop–drop coalescence in the systems studied by Moens et al. [96].

The effect of the cooling rate on partial coalescence also remains controversial. At the low cooling rate, the crystal growth process is favored over the nucleation process, resulting in a smaller number of larger-in-size crystals. In contrast, at higher cooling rates, the nucleation process prevails, and numerous smaller crystallites are formed [94,96]. Fredrick et al. [94] hypothesized that the larger crystals protrude over a longer distance into the aqueous phase, therefore enhancing the partial coalescence. Degner et al. [97] also concluded that slow freezing enhances the drop–drop aggregation. However, Moens et al. [96,98], in their studies, reported the opposite effect of the cooling rate — the emulsions were more stable at slower cooling rates, under otherwise equivalent conditions.

These apparent discrepancies could be resolved by considering the fact that depending on the surfactant used for emulsion stabilization, fat crystallization may nucleate at two different locations — in the drop interior or on the drop surface. Indeed, several studies revealed that partial coalescence depends strongly on the melting temperature of the low-molecular-mass surfactants present [19,20,94,99]. The long-chain saturated monoglycerides form adsorption layers, which may freeze before the freezing of the oil phase inside the drops, whereas the surfactants containing double bonds (oleates) form fluid adsorption layers, which cannot crystallize prior to the crystallization of the drop interior.

Thus, we see that the effects of freezing of the surfactant adsorption layer and the rate of emulsion cooling are interrelated. For fluid adsorption layers, drop freezing nucleates in the drop interior, and the fat crystals grow toward the drop periphery. On slow cooling, larger crystallites are formed, which have greater protrusion into the aqueous phase, thus enhancing the partial coalescence (cf Figure 5a,b) [99].

In contrast, for presolidified adsorption layers, drop freezing starts from the drop surface and evolves

toward the drop interior, templating lamella planes of the fat crystals with an orientation parallel to the oil–water interface [20,95]. Here, slow cooling should create a fewer number of surface nuclei, which slowly grow toward the drop interior, without breaking the neighboring emulsion films (Figure 5d). Fast cooling, however, should lead to dense coverage of the drop surface with a fragile crust of small crystallites, which may become corrugated with the subsequent freezing and shrinking of the remaining oil inside the droplets, thus inducing partial coalescence of the neighboring drops (Figure 5e).

In other words, the effect of the cooling rate could be opposite, depending on the type of the surfactant used and on the ensuing location of fat crystal nucleation, thus explaining the apparent discrepancies reported in the literature.

Interfacial nucleation was also related to the observed increased stability toward partial coalescence in paraffin-in-water emulsions, studied as phase change materials for thermal storage [19,21]. In these systems, the emulsifiers, which are in a liquid state around the temperatures of paraffin crystallization, were found to promote partial coalescence [19,20].

We conclude that surface freezing and the related surface nucleation of fat/alkane crystals change dramatically the behavior of the emulsions in the process of partial coalescence. Further studies are needed to build

up a univocal mechanistic explanation of the wide variety of behaviors observed and to reveal the convenient factors for process control.

### Surface freezing in drop self-shaping and drop-bursting phenomena

The crystallization of surfactant adsorption layers on the surface of oil drops turned out to be a crucial step in two other, recently discovered phenomena, observed in cooled emulsions — drop self-shaping [100\*–102] and drop self-emulsification [103\*,104] (Figure 6). The main observations and the role of surface freezing in these phenomena are discussed briefly in this section.

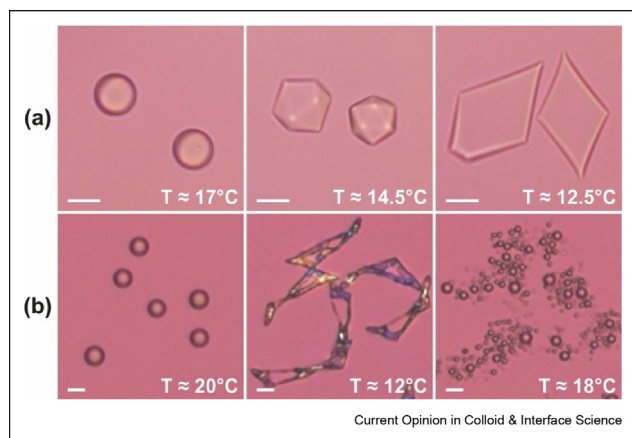
Emulsion drops from various long-chain organic substances, such as alkanes, alkenes, alcohols, and their mixtures, were observed to exhibit a spectacular series of shape transformations on emulsion cooling, but only when the oil drops are stabilized by surfactants with long saturated hydrophobic chains, for example, with 16 or 18 carbon atoms [100\*,101]. Two different mechanisms were proposed in the literature and are currently debated to explain this phenomenon, both of them starting with freezing of the surfactant adsorption layer on the oil drop surface as a first step triggering a series of subsequent steps.

Denkov *et al.* [100\*] assumed that the frozen surfactant layer serves as a template for ordering of the neighboring oil molecules, next to the drop surface, into molecular multilayers of the so-called ‘rotator’ or ‘plastic’ phases [105\*\*,106]. These rotator phases have certain viscoelasticity that may overcome the capillary pressure of the droplets, which acts to preserve the spherical drop shape. Differential scanning calorimetry was used to determine the thickness of the formed rotator phases, which was found to vary between 2 and ca. 80 molecular layers, depending on the specific emulsion composition and drop size [106]. On the basis of this mechanism, a detailed theoretical model was proposed that explained the experimentally observed sequence of drop transformations [107,108].

In an alternative explanation, Guttman *et al.* [109\*,110] proposed that the frozen surfactant monolayer could (*per se*) drive the drop-shape transformations owing to ultralow and even ‘transiently negative’ interfacial tension, which may occur in some nonequilibrium systems. Indeed, the Krafft point of the C<sub>18</sub>TAB surfactant, used in the experiments of Guttman *et al.* [109\*,110], is  $\approx 36^\circ\text{C}$  [111], which suggests that the experiments with hexadecane drops (melting temperature  $\approx 18^\circ\text{C}$ ) are performed at temperatures lower than the surfactant Krafft point, that is, under nonequilibrium conditions.

It was shown later by these two groups that the shaped drops could be polymerized under appropriate

Figure 6



Microscopy images showing: **(a)** drop self-shaping and **(b)** drop-bursting phenomena in cooled emulsions. **(a)** Hexadecane droplets, dispersed in 1.5 wt% aqueous solution of Tween 40, have initially the characteristic spherical shape (left). On cooling, the drops transform into regular polyhedrons (in the middle) that gradually flatten until tetragonal platelets are formed (right). **(b)** From the initial six spherical hexadecane droplets (left), six frozen platelets are formed on cooling (in the middle). On subsequent heating, the melting platelets disperse spontaneously into hundreds of smaller droplets (right). The hexadecane emulsion in **(b)** is stabilized by 1.5 wt% Brij S20 nonionic surfactant. In all images, the scale bars are 10  $\mu\text{m}$ .

conditions and that even submicrometer droplets can undergo self-shaping on emulsion cooling [112,113].

The current brief review is not a place to debate about the different possible mechanisms of drop self-shaping. Here we just note that the interfacial tension was measured to be in the range between ca. 2 mN/m and 10 mN/m, viz, it was not ultralow or negative for the surfactant–oil systems, studied by Denkov et al. [100–106]. Therefore, the mechanism, proposed by Guttman et al. [109\*,110], cannot explain the results reported in the studies by Denkov et al [100\*] and Cholakova et al [101], although it might be operative for the experimental systems described in the studies by Guttman et al [109\*,110]. Note that the two research groups always worked with different surfactant–oil systems and one cannot exclude the possibility that different mechanisms were operative in their studies.

In related studies, a drop ‘self-emulsification’ phenomenon was observed on cooling and heating of emulsions, stabilized by long-chain surfactants [103\*,104]. Several detailed mechanisms were observed by optical microscopy to induce spontaneous drop fragmentation. All these mechanisms were related to the formation of dense adsorption layers which freeze on emulsion cooling before the complete solidification of the oil drops. Some of these mechanisms are extremely efficient — after freezing and on melting, the initial drops may burst into hundreds and thousands of submicrometer droplets without any mechanical input to the emulsion [103\*,104].

Related to these phenomena could be the study by Kovacik et al. [114] who performed temperature-dependent second-harmonic sum-frequency and light scattering measurements on alkane-in-water nano-emulsions, stabilized by  $C_n$ TAB. These authors concluded that, when the surfactant tail is longer than the alkane chain length, a transformation in the interfacial water ordering and freezing of the drop surface layer occur first, followed by solidification of the drop interior. It was not clarified whether the nanodroplets in the study by Kovacik et al [114] changed their shape when long-chain  $C_n$ TAB surfactants were used for emulsion stabilization — a possibility which is worthy to be explored in further studies.

### Saponin-stabilized emulsions

Saponin-stabilized emulsions were recently studied by several groups. In the study by Ralla et al [115], a very good thermal stability of medium-chain triglyceride miglyol-in-water emulsions, stabilized by the *Yucca* saponin extract, was reported in the pH range between 5 and 9. In a separate study, the emulsions stabilized by QSs were found to be sensitive to the variations in pH and electrolyte concentration [116,117]. In both cases,

the emulsion stability was explained with electrostatic repulsion between the oil drops, without invoking any effects related to the surface viscoelasticity of the saponin adsorption layers. On the other hand, the studies of such layers at the oil–water interfaces showed very high surface viscoelasticity for some saponin–oil combinations [64]. Peculiar in shape, nonspherical emulsion drops were observed in the respective emulsions [64]. The latter observations imply that the properties of some saponin-stabilized emulsions could be governed by the surface viscoelasticity of the respective adsorption layers — a possibility which should be checked in the future.

In a very interesting study [118\*], Chen et al. developed a procedure for formation of concentrated nano-emulsion of droplets with diameter of  $\approx 100$  nm, stabilized by QSs. These nanoemulsions were aerated to obtain stable and highly viscoelastic foams with a hierarchical internal structure. The peculiar properties of this system were explained with the high surface viscoelasticity of the saponin adsorption layers and with the strong adhesion of the nanodroplets to the bubble surfaces, which resulted in Pickering-type foam stabilization.

Similarly, aerated emulsions were obtained from GA (another type of saponin) [119]. Nanofibrils and the fibrillary network (hydrogel) were formed by GA in the aqueous phase, which blocked the Plateau channels and stabilized the foams at low temperature. The saponin hydrogel was destroyed at higher temperature, thus deteriorating foam stability.

The aforementioned overview evidences that further studies are needed to deepen our understanding of the factors controlling the surface phase transition in emulsions and the related complex phenomena.

### Outlook

The phase transitions on the surface of bubbles have been used for years to control dynamic properties of foams. Still, very little is known about the detailed structure of the adsorption layers in the most widely used surfactant–cosurfactant and saponin-containing systems with high surface viscoelasticity. Therefore, one may expect that the established experimental methods for surface structural analysis will be used in the next years to understand better the molecular structure–function relations in the formation of viscoelastic adsorption layers. The computer methods of molecular modeling (e.g. molecular dynamics) will be also a very useful tool to analyze the role and relative contribution of the various types of intermolecular forces in the viscoelastic response of surfactant adsorption layers. Deeper understanding would allow the researchers and practitioners to design in a more rational

way specific surfactant mixtures with desired surface properties and controlled response to variations of pH, temperature, electrolytes, and other stimuli. Interesting new approaches for foam control emerged in the last years, such as the use of photosensitive surfactants and fluorinated surfactants, both of which exhibit specific properties, very different from those of the known conventional surfactants.

The approaches for control of the surface phase transitions in emulsions are even less understood. Only recently, the accumulated knowledge about surface freezing of surfactant adsorption layers at the oil–water interfaces has been invoked to control the stability of regular emulsions. This area will certainly expand in the coming years owing to the importance of emulsion partial coalescence in several industrial areas and to the discovery of the new intriguing phenomena of drop self-shaping and drop bursting in cooled emulsions, all strongly dependent on the process of surface freezing. Other new and unexpected phenomena could emerge when more intensive and systematic studies are performed on the role of surface phase transitions in these systems.

As evidenced in various studies, the surface phase transitions could trigger and could be involved in a complex interplay with other aggregation phenomena, such as surface templating and precipitation, surfactant–polymer complexation, and surfactant modification of particle hydrophobicity with subsequent surface or bulk particle aggregation/gelation. Such combined mechanisms were found to create ultrastable foams and emulsions, with various potential applications, including their use as precursors for the fabrication of porous materials with a hierarchical structure. The role of surface phase transitions has been recognized in these complex phenomena but has not been systematically studied yet. Therefore, understanding the role of surface phase transitions and revealing the main factors for their control would enhance our approaches for rational control over these complex phenomena with many potential applications.

## Conflict of interest statement

Nothing declared.

## Acknowledgements

This work was supported by the Bulgarian Ministry of Education and Science, under the National Research Programme “Healthy Foods for a Strong Bio-Economy and Quality of Life”, approved by DCM #577/17.08.2018. The study falls under the umbrella of COST action CA17120 ‘Chemobionics,’ funded by the program Horizon 2020 of the EU.

## References

Papers of particular interest, published within the period of review, have been highlighted as:

- \* of special interest
- \*\* of outstanding interest

1. Miles GD, Ross J, Shedlovsky L: **Film drainage: a study of the flow properties of films of solutions of detergents and the effect of added materials.** *J Am Oil Chem Soc* 1950, **27**: 268–273, <https://doi.org/10.1007/BF02634904>.
2. Mysels KJ, Shinoda K, Frankel S: **Soap films: studies of their thinning.** New York: Pergamon Press; 1959.
3. Epstein MB, Wilson A, Jakob CW, Conroy LE, Ross J: **Film drainage transition temperatures and phase relations in the system sodium lauryl sulfate, lauryl alcohol and water.** *J Phys Chem* 1954, **58**:860–864, <https://doi.org/10.1021/j150520a014>.
4. Ivanov IB, Dimitrov DS: **Thin film drainage.** In *Thin liquid films.* Edited by Ivanov IB, New York: M. Dekker; 1988.
5. Langevin D: **Rheology of adsorbed surfactant monolayers at fluid surfaces.** *Annu Rev Fluid Mech* 2014, **46**:47–65, <https://doi.org/10.1146/annurev-fluid-010313-141403>.
6. Joye J-L, Hirasaki GJ, Miller CA: **Asymmetric drainage in foam films.** *Langmuir* 1994, **10**:3174–3179, <https://doi.org/10.1021/la00021a046>.
7. Bergeron V: **Disjoining pressures and film stability of alkyl-trimethylammonium bromide foam films.** *Langmuir* 1997, **13**: 3474–3482, <https://doi.org/10.1021/la970004q>.
8. Weaire D, Hutzler S, Verbist G, Peters E: **A review of foam drainage.** *Adv Chem Phys* 1997, **102**:315–374, <https://doi.org/10.1002/9780470141618.ch5>.
9. Koehler SA, Hilgenfeldt S, Stone HA: **A generalized view of foam drainage: experiment and theory.** *Langmuir* 2000, **16**: 6237–6341, <https://doi.org/10.1021/la9913147>.
10. Stoyanov S, Dushkin C, Langevin D, Weaire D, Verbist G: **Effect of the rheology on foam drainage.** *Langmuir* 1998, **14**: 4663–4665, <https://doi.org/10.1021/la9713356>.
11. Durand M, Martinot G, Langevin D: **Liquid flow through aqueous foams: from the plateau border-dominated regime to the node-dominated regime.** *Phys Rev E* 1999, **60**: R6307–R6308, <https://doi.org/10.1103/physreve.60.r6307>.
12. Durand M, Langevin D: **Physicochemical approach to the theory of foam drainage.** *Eur Phys J E* 2002, **7**:35–44, <https://doi.org/10.1140/epje/i200101092>.
13. Denkov ND, Subramanian V, Gurovich D, Lips A: **Wall slip and viscous dissipation in sheared foams: effect of surface mobility.** *Colloids Surf, A* 2005, **263**:129–145, <https://doi.org/10.1016/j.colsurfa.2005.02.038>.
14. Denkov ND, Tcholakova S, Golemanov K, Ananthapadmanabhan KP, Lips A: **The role of surfactant type and bubble surface mobility in foam rheology.** *Soft Matter* 2009, **5**:3389–3408, <https://doi.org/10.1039/b903586a>.
15. Mitrinova Z, Tcholakova S, Golemanov K, Denkov N, Vethamuthu M, Ananthapadmanabhan KP: **Surface and foam properties of SLES + CAPB + fatty acid mixtures: effect of pH for C12-C16 acids.** *Colloids Surf, A* 2013, **438**:186–198, <https://doi.org/10.1016/j.colsurfa.2012.12.011>.
16. Stocco A, Carriere D, Cottat M, Langevin D: **Interfacial behavior of catanionic surfactants.** *Langmuir* 2010, **26**:10663–10669, <https://doi.org/10.1021/la100954v>.
17. Varade D, Carriere D, Arriaga LR, Fameau A-L, Rio E, Langevin D, Drenckhan W: **On the origin of the stability of foams made from catanionic surfactant mixtures.** *Soft Matter* 2011, **7**:6557–6570, <https://doi.org/10.1039/c1sm05374d>.
18. Tcholakova S, Mitrinova Z, Golemanov K, Denkov ND, Vethamuthu M, Ananthapadmanabhan KP: **Control of Ostwald ripening by using surfactants with high surface modulus.** *Langmuir* 2011, **27**:14807–14819, <https://doi.org/10.1021/la203952p>.
19. Golemanov K, Tcholakova S, Denkov N, Gurkov T: **Selection of surfactants for stable paraffin-in-water dispersions, undergoing solid-liquid transition of the dispersed particles.** *Langmuir* 2006, **22**:3560–3569, <https://doi.org/10.1021/la053059y>.

20. Goibier L, Lecomte S, Leal-Calderon F, Faure C: **The effect of surfactant crystallization on partial coalescence in O/W emulsions.** *J Colloid Interface Sci* 2017, **500**:304–314, <https://doi.org/10.1016/j.jcis.2017.04.021>.
21. Shchukina EM, Graham M, Zheng Z, Shchukin DG: **Nano-encapsulation of phase change materials for advanced thermal energy storage systems.** *Chem Soc Rev* 2018, **47**: 4156–4175, <https://doi.org/10.1039/c8cs00099a>.
22. Vollhardt D, Fainerman VB: **Characterisation of phase transition in adsorbed monolayers at the air/water interface.** *Adv Colloid Interface Sci* 2010, **154**:1–19, <https://doi.org/10.1016/j.cis.2010.01.003>.
23. Gaines GL: *Insoluble monolayers at the liquid-gas interface.* New York: Wiley-Interscience; 1966.
24. Adamson AW, Gast AP: *Physical chemistry of surfaces.* 6th ed. New Jersey: Wiley; 1997.
25. Lautz C, Fischer TM, Weygand M, Lösche M, Howes PB, Kjaer K: **Determination of alkyl chain tilt angles in Langmuir monolayers: a comparison of Brewster angle autocorrelation spectroscopy and x-ray diffraction.** *J Chem Phys* 1998, **108**: 4640–4646, <https://doi.org/10.1063/1.475874>.
26. Henon S, Meunier J: **Microscope at the Brewster angle: direct observation of first order phase transitions in monolayers.** *Rev Sci Instrum* 1991, **62**:936–939, <https://doi.org/10.1063/1.1142032>.
27. Möhwald H: **Direct characterization of monolayers at the air-water interface.** *Thin Solid Films* 1988, **159**:1–15, [https://doi.org/10.1016/0040-6090\(88\)90612-8](https://doi.org/10.1016/0040-6090(88)90612-8).
28. Als-Nielsen J, Jacquemain D, Kjaer K, Leveiller F, Lahav M, Leiserowitz L: **Principles and applications of grazing incidence X-ray and neutron scattering from ordered molecular monolayers at the air-water interface.** *Phys Rep* 1994, **246**: 251–313, [https://doi.org/10.1016/0370-1573\(94\)90046-9](https://doi.org/10.1016/0370-1573(94)90046-9).
29. Schwartz DK: **Langmuir-Blodgett film structure.** *Surf Sci Rep* 1997, **27**:241–334, [https://doi.org/10.1016/S0167-5729\(97\)00003-4](https://doi.org/10.1016/S0167-5729(97)00003-4).
30. Russev SC, Alexandrov N, Marinova KG, Danov KD, Denkov ND, Lyutov L, Vulchev V, Bilke-Krause C: **Instrument and methods for surface dilatational rheology measurements.** *Rev Sci Instrum* 2008, **79**:104102, <https://doi.org/10.1063/1.3000569>.
31. Alexandrov N, Marinova KG, Danov KD, Ivanov IB: **Surface dilatational rheology measurements for oil-water systems with viscous oils.** *J Colloid Interface Sci* 2009, **339**:545–550, <https://doi.org/10.1016/j.jcis.2009.08.002>.
32. Brooks CF, Fuller GG, Frank CW, Robertson CR: **An interfacial stress rheometer to study rheological transitions in monolayers at the air-water interface.** *Langmuir* 1999, **15**: 2450–2459, <https://doi.org/10.1021/la980465r>.
33. Reynaert S, Brooks CF, Moldenaers P, Vermant J, Fuller GG: **Analysis of the magnetic rod interfacial stress rheometer.** *J Rheol* 2008, **52**:261–285, <https://doi.org/10.1122/1.2798238>.
34. Vandebriel S, Franck A, Fuller GG, Moldenaers P, Vermant J: **A double wall-ring geometry for interfacial shear rheometry.** *Rheol Acta* 2010, **49**:131–144, <https://doi.org/10.1007/s00397-009-0407-3>.
35. Sachan AK, Choi SQ, Kim KH, Tang Q, Hwang L, Lee KYC, Squires TM, Zasadzinski JA: **Interfacial rheology of coexisting solid and fluid monolayers.** *Soft Matter* 2017, **13**:1481–1492, <https://doi.org/10.1039/c6sm02797k>.
- The authors show that monolayer fluidity may vary by orders of magnitude upon small changes in the monolayer composition and that the formation of surface condensed phase increases dramatically the overall monolayer viscosity.
36. Langevin D, Monroy F: **Marangoni stresses and surface compression rheology of surfactant solutions. Achievements and problems.** *Adv Colloid Interface Sci* 2014, **206**:141–149, <https://doi.org/10.1016/j.cis.2014.01.006>.
37. Langevin D: **Surface shear rheology of monolayers at the surface of water.** *Adv Colloid Interface Sci* 2014, **207**:121–130, <https://doi.org/10.1016/j.cis.2013.10.030>.
38. Zuo YY, Chen R, Wang X, Yang J, Policova Z, Neumann AW: **Phase transitions in dipalmitoylphosphatidylcholine monolayers.** *Langmuir* 2016, **32**:8501–8506, <https://doi.org/10.1021/acs.langmuir.6b01482>.
39. Xu L, Zuo YY: **Reversible transitions in the phospholipid monolayer.** *Langmuir* 2018, **34**:8694–8700, <https://doi.org/10.1021/acs.langmuir.8b01544>.
40. Cuenca VE, Leyes MF, Falcone RD, Correa NM, Langevin D, Ritacco H: **Interfacial dynamics and its relation with “negative” surface viscosities measured at water-air interfaces covered with a cationic surfactants.** *Langmuir* 2019, **35**: 8333–8343, <https://doi.org/10.1021/acs.langmuir.9b00534>.
41. Golemanov K, Denkov ND, Tcholakova S, Vethamuthu M, Lips A: **Surfactant mixtures for control of bubble surface mobility in foam studies.** *Langmuir* 2008, **24**:9956–9961, <https://doi.org/10.1021/la8015386>.
42. Wang J, Nguyen AV, Farrokhpay S: **Effects of surface rheology and surface potential on foam stability.** *Colloids Surf, A* 2016, **488**:70–81, <https://doi.org/10.1016/j.colsurfa.2015.10.016>.
43. Mitrinova Z, Tcholakova S, Popova Z, Denkov N, Dasgupta BR, Ananthapadmanabhan KP: **Efficient control of the rheological and surface properties of surfactant solutions containing C8-C18 fatty acids as cosurfactants.** *Langmuir* 2013, **29**: 8255–8265, <https://doi.org/10.1021/la401291a>.
- Systematic study on the role of fatty acid chain length on the bulk and surface properties of surfactant solutions. Optimal chain-length of C14 and C16 is found to create viscoelastic adsorption layers with very high surface modulus.
44. Arnold A, Cousin F, Salonen A, Jalmes AS, Perez AA, Fameau A-L: **Controlling foam stability with the ratio of myristic acid to choline hydroxide.** *Langmuir* 2018, **34**:11076–11085, <https://doi.org/10.1021/acs.langmuir.8b02261>.
- Maximum foam stability is observed at pH around the pK of myristic acid which is explained with the formation of mixed adsorption layer of charged and non-charged surfactant species. The choline is used as a bulky counterion to increase the myristic acid solubility at low pH.
45. Small DM: *The physical chemistry of lipids. From alkanes to phospholipids.* New York: Plenum; 1986.
46. Marinova KG, Naydenova KT, Basheva ES, Bauer F, Tropsch J, Franke J: **New surfactant mixtures for fine foams with slowed drainage.** *Colloids Surf, A* 2017, **523**:54–61, <https://doi.org/10.1016/j.colsurfa.2017.03.050>.
47. Wang Y, Ge J, Song K, Zhang G, Jiang P: **Effect of organic acid chain length on foam performance in a porous medium.** *J Disp Sci Technol* 2017, **38**:1448–1458, <https://doi.org/10.1080/01932691.2016.1251324>.
48. Mensire R, Wexler JS, Guibaud A, Lorenceau EF, Stone HA: **Surfactant and aqueous foam-driven oil extraction from micropatterned surfaces.** *Langmuir* 2016, **32**:13149–13158, <https://doi.org/10.1021/acs.langmuir.6b03225>.
49. Wang Y, Ge J, Zhang G, Jiang P: **Effect of organic acid on lauroamide propyl betaine surface dilatational modulus and foam performance in a porous medium.** *J Disp Sci Technol* 2017, **38**:1348–1354, <https://doi.org/10.1080/01932691.2016.1183210>.
50. Wang Y, Ge J, Zhang G, Jiang P, Song K, Zhang W: **Effect of surface dilatational modulus on foam flow in a porous medium, Tenside, Surfactants.** *Detergents* 2017, **54**:327–333, <https://doi.org/10.3139/113.110508>.
51. Dollet B, Bocher C: **Flow of foam through a convergent channel.** *Eur Phys J E* 2015, **38**:123, <https://doi.org/10.1140/epje/i2015-15123-3>.
52. Vitry Y, Dorbolo S, Vermant J, Scheid B: **Controlling the lifetime of antibubbles.** *Adv Colloid Interface Sci* 2019, **270**:73–86, <https://doi.org/10.1016/j.cis.2019.05.007>.
53. Arriaga LR, Varade D, Carriere D, Drenckhan W, Langevin D: **Adsorption, organization, and rheology of catanionic layers at the air/water interface.** *Langmuir* 2013, **29**:3214–3222, <https://doi.org/10.1021/la304868n>.
- The adsorption of catanionic complexes of myristic acid + CTAB is studied. LC domains in LE matrix are observed in the adsorption

monolayers. These layers behave as soft glassy materials and melt above a certain temperature.

54. Rio E, Drenckhan W, Salonen A, Langevin D: **Unusually stable liquid foams.** *Adv Colloid Interface Sci* 2014, **205**:74–86, <https://doi.org/10.1016/j.cis.2013.10.023>.

The approaches to obtain ultrastable foams are reviewed. Special emphasis is given on the coupling between the three main modes of foam destabilization (bubble coarsening, liquid drainage, bubble coalescence) and on the strategies for the simultaneous arrest of them all.

55. Zhang L, Mikhailovskaya A, Yazhgur P, Muller F, Cousin F, Langevin D, Wang N, Salonen A: **Precipitating sodium dodecyl sulfate to create ultrastable and stimulative foams.** *Angew Chem Int Ed* 2015, **54**:9533–9536, <https://doi.org/10.1002/anie.2015.03236>.

56. Lyadinskaya VV, Lin S-Y, Michailov AV, Povolotskiy AV, Noskov BA: **Phase transitions in DNA/surfactant adsorption layers.** *Langmuir* 2016, **32**:13435–13445, <https://doi.org/10.1021/acs.langmuir.6b03396>.

A variety of intriguing phenomena are observed in mixed adsorption layers of DNA molecules with DTAB or CTAB surfactants. Several experimental methods are used to reveal the structure of the formed mixed layers and to explain the main observations.

57. Xu H, Li P, Ma K, Welbourn RJL, Penfold J, Thomas RK, Roberts DW, Petkov JT: **The role of competitive counterion adsorption on the electrolyte induced surface ordering in methyl ester sulfonate surfactants at the air-water interface.** *J Colloid Interface Sci* 2019, **533**:154–160, <https://doi.org/10.1016/j.jcis.2018.08.061>.

58. Xu H, Thomas RK, Penfold J, Li PX, Ma K, Welbourne RJL, Roberts DW, Petkov JT: **The impact of electrolyte on the adsorption of the anionic surfactant methyl ester sulfonate at the air-solution interface: surface multilayer formation.** *J Colloid Interface Sci* 2018, **512**:231–238, <https://doi.org/10.1016/j.jcis.2017.10.064>.

59. Xu H, Li P, Ma K, Welbourn RJL, Douth J, Penfold J, Thomas RK, Roberts DW, Petkov JT, Choo KL, Khoo SY: **Adsorption and self-assembly in methyl ester sulfonate surfactants, their eutectic mixtures and the role of electrolyte.** *J Colloid Interface Sci* 2018, **516**:456–465, <https://doi.org/10.1016/j.jcis.2018.01.086>.

60. Xu H, Penfold J, Thomas RK, Petkov JT, Tucker I, Webster JRP, Grillo I, Terry A: **Ion specific effects in trivalent counterion induced surface and solution self-assembly of the anionic surfactant sodium polyethylene glycol monododecyl ether sulfate.** *Langmuir* 2014, **30**:4694–4702, <https://doi.org/10.1021/la500937g>.

61. Pagureva N, Tcholakova S, Golemanov K, Denkov N, Pelan E, Stoyanov SD: **Surface properties of adsorption layers formed from triterpenoid and steroid saponins.** *Colloids Surf, A* 2016, **491**:18–28, <https://doi.org/10.1016/j.colsurfa.2015.12.001>.

The surface properties of several saponins are analyzed to show that the high surface viscoelasticity, observed for some of them, is related to a very strong attraction between the saponin molecules in their adsorption layers.

62. Tsihranska S, Ivanova A, Tcholakova S, Denkov N: **Self-assembly of escin molecules at the air-water interface as studied by molecular dynamics.** *Langmuir* 2017, **33**:8330–8341, <https://doi.org/10.1021/acs.langmuir.7b01719>.

Using molecular dynamic simulations the authors show that the strong attraction between the saponin molecules in adsorption layers is due to the combined effect of long-range dipole–dipole interactions and short-range hydrogen bonds.

63. Stanimirova R, Marinova K, Tcholakova S, Denkov ND, Stoyanov S, Pelan E: **Surface rheology of saponin adsorption layers.** *Langmuir* 2011, **27**:12486–12498, <https://doi.org/10.1021/la202860u>.

64. Golemanov K, Tcholakova S, Denkov N, Pelan E, Stoyanov SD: **The role of the hydrophobic phase in the unique rheological properties of saponin adsorption layers.** *Soft Matter* 2014, **10**:7034–7044, <https://doi.org/10.1039/c4sm00406j>.

65. Wojciechowski K: **Surface activity of saponin from *Quillaja* bark at the air/water and oil/water interfaces.** *Colloids Surf, B* 2013, **108**:95–102, <https://doi.org/10.1016/j.colsurfb.2013.02.008>.

66. Golemanov K, Tcholakova S, Denkov N, Pelan E, Stoyanov SD: **Remarkably high surface visco-elasticity of adsorption layers of triterpenoid saponins.** *Soft Matter* 2013, **9**:5738–5752, <https://doi.org/10.1039/c3sm27950b>.

67. Kezwon A, Wojciechowski K: **Interaction of *Quillaja* bark saponins with food-relevant proteins.** *Adv Colloid Interface Sci* 2014, **209**:185–195, <https://doi.org/10.1016/j.cis.2014.04.005>.

Strong surface complexation in mixed adsorption layers of *Quillaja* saponin (QS) with proteins and lipids is reported.

68. Böttcher S, Scampicchio M, Drusch S: **Mixtures of saponins and beta-lactoglobulin differ from classical protein/surfactant-systems at the air-water interface.** *Colloids Surf, A* 2016, **506**:765–773, <https://doi.org/10.1016/j.colsurfa.2016.07.057>.

69. Ulaganathan V, Castillo LD, Webber JL, Ho TTM, Ferri JK, Krasowska M, Beattie DA: **The influence of pH on the interfacial behaviour of *Quillaja* bark saponin at the air-solution interface.** *Colloids Surf, B* 2019, **176**:412–419, <https://doi.org/10.1016/j.colsurfb.2019.01.017>.

70. Böttcher S, Drusch S: **Interfacial properties of saponin extracts and their impact on foam characteristics.** *Food Biophys* 2016, **11**:91–100, <https://doi.org/10.1007/s11483-015-9420-5>.

71. Böttcher S, Drusch S: **Saponins – self-assembly and behavior at aqueous interfaces.** *Adv Colloid Interface Sci* 2017, **243**:105–113, <https://doi.org/10.1016/j.cis.2017.02.008>.

72. Santini E, Jarek E, Ravera F, Liggieri L, Warszynski P, Krzan M: **Surface properties and foamability of saponin and saponin-chitosan systems.** *Colloids Surf, B* 2019, **181**:198–206, <https://doi.org/10.1016/j.colsurfb.2019.05.035>.

73. Müller LE, Schiedeck G: **Physical properties of botanical surfactants.** *Sci Total Environ* 2018, **610**:11133–1137, <https://doi.org/10.1016/j.scitotenv.2017.08.193>.

74. Ma L, Li Q, Du Z, Su E, Liu X, Wan Z, Yang X: **A natural supramolecular saponin hydrogelator for creation of ultrastable and thermostimulable food-grade foams.** *Adv Mater Interfaces* 2019, **6**:1900417, <https://doi.org/10.1002/admi.201900417>.

75. Kovalenko A, Polavarapu P, Gallani J-L, Pourroy G, Waton G, Krafft MP: **Super-elastic air/water interfacial films self-assembled from soluble surfactants.** *Chem PhysChem* 2014, **15**:2440–2444, <https://doi.org/10.1002/cphc.201402248>.

Extremely high surface viscoelasticity, up to 900 mN/m, is measured with solutions of fluorinated surfactants of moderate chain-length (8–10 C atoms). The forces which lead to condensation of the respective adsorption layer need to be better understood.

76. Noskov B, Akentiev AV, Lin S-Y: **Dynamic properties of adsorption layers of heptadecafluoro-1-nonanol. Effect of surface phase transitions.** *J Mol Liq* 2019, **282**:316–322, <https://doi.org/10.1016/j.molliq.2019.03.009>.

Another study which shows very high surface viscoelasticity, up to 250 mN/m, for solutions of fluorinated surfactants with moderate chain-length.

77. Jiang N, Sheng Y, Li C, Li S: **Surface activity, foam properties and aggregation behavior of mixtures of short-chain fluorocarbon and hydrocarbon surfactants.** *J Mol Liq* 2018, **268**:149–255, <https://doi.org/10.1016/j.molliq.2018.07.055>.

78. Eastoe J, Vesperinas A: **Self-assembly of light-sensitive surfactants.** *Soft Matter* 2005, **1**:338–347, <https://doi.org/10.1039/b510877m>.

79. Chevallier E, Mamane A, Stone HA, Tribet C, Lequeux F, Monteux C: **Pumping-out photo-surfactants from an air–water interface using light.** *Soft Matter* 2011, **7**:7866–7874, <https://doi.org/10.1039/c1sm05378g>.

80. Chevallier E, Monteux C, Lequeux F, Tribet C: **Photofoams: remote control of foam destabilization by exposure to light using an azobenzene surfactants.** *Langmuir* 2012, **28**:2308–2312, <https://doi.org/10.1021/la204200z>.

81. Chevallier E, Saint-Jalmes A, Cantat I, Lequeux F, Monteux C: **Light induced flows opposing drainage in foams and thin-films using photosurfactants.** *Soft Matter* 2013, **9**:7054–7060, <https://doi.org/10.1039/C3SM50258A>.

This and the previous studies by the same group report and explain several unexpected phenomena, all of them caused by non-trivial changes in the surface and bulk properties of the surfactant molecules after their cis–trans isomerization upon light illumination.

82. Jiang J, Ma Y, Cui Z: **Smart foams based on dual stimuli-responsive surfactant.** *Colloids Surf, A* 2017, **513**:287–291, <https://doi.org/10.1016/j.colsurfa.2016.10.056>.
83. Lei Q, Bain CD: **Surfactant-induced surface freezing at the alkane-water interface.** *Phys Rev Lett* 2004, **92**:176103, <https://doi.org/10.1103/PhysRevLett.92.176103>.
84. Wilkinson KM, Qunfang L, Bain CD: **Freezing transitions in mixed surfactant/alkane monolayers at the air-solution interface.** *Soft Matter* 2006, **2**:66–76, <https://doi.org/10.1039/b511561b>.
85. Sloutskin E, Sapir Z, Tamam L, Ocko BM, Bain CD, Deutsch M: **Freezing transition of Langmuir-Gibbs alkane films on water.** *Thin Solid Films* 2007, **515**:5664–5668, <https://doi.org/10.1016/j.tsf.2006.12.007>.
86. Yefet S, Sloutskin E, Tamam L, Sapir Z, Deutsch M, Ocko BM: **Surfactant-induced phases in water-supported alkane monolayers: II. Structure.** *Langmuir* 2014, **30**:8010–8019, <https://doi.org/10.1021/la501589t>.  
The structure of the surface phases in adsorption layers of  $C_m$ TAB surfactants, in the presence of alkanes of different chain-lengths,  $C_nH_{2n+2}$ , is studied. Several monolayer and bilayer structures are observed, depending on the difference between  $n$  and  $m$ .
87. Matsubara H, Shigeta T, Takata Y, Ikeda N, Sakamoto H, Takiue T, Aratono M: **Effect of molecular structure of oil on wetting transition on surfactant solutions.** *Colloids Surf, A* 2007, **301**:141–146, <https://doi.org/10.1016/j.colsurfa.2006.12.040>.
88. Tokiwa Y, Sakamoto H, Takiue T, Aratono M, Matsubara H: **Effect of alkane chain length and counterion on the freezing transition of cationic surfactant adsorbed film at alkane mixture-water interfaces.** *J Phys Chem, B* 2015, **119**:6235–6241, <https://doi.org/10.1021/acs.jpcc.5b02448>.
89. Matsubara H, Aratono M: **Unique interfacial phenomena on macroscopic and colloidal scales induced by two-dimensional phase transitions.** *Langmuir* 2019, **35**:1989–2001, <https://doi.org/10.1021/acs.langmuir.8b01203>.  
Very comprehensive and detailed review on the variety of interfacial phenomena, related to the surface phase transitions.
90. Tokiwa Y, Sakamoto H, Takiue T, Aratono M, Matsubara H: **Effect of surface freezing on stability of oil-in-water emulsions.** *Langmuir* 2018, **34**:6205–6209, <https://doi.org/10.1021/acs.langmuir.8b01088>.  
Study relating directly the phase transition in the adsorption layer on the surface of oil drops to the coalescence stability of the drops in the respective oil-in-water emulsions. The emulsion stability is found to decrease significantly after melting the surfactant adsorption layer.
91. McClements DJ: *Food emulsions: principles, practices, and techniques*. 3rd ed. Boca Raton: CRC Press; 2015.
92. Boode K, Walstra P: **Partial coalescence in oil-in-water emulsions 1. Nature of the aggregation.** *Colloids Surf, A* 1993, **81**:121–137, [https://doi.org/10.1016/0927-7757\(93\)80239-B](https://doi.org/10.1016/0927-7757(93)80239-B).
93. Boode K, Walstra P, de Groot-Mostert AEA: **Partial coalescence in oil-in-water emulsions 2. Influence of the properties of the fat.** *Colloids Surf, A* 1993, **81**:139–151, [https://doi.org/10.1016/0927-7757\(93\)80240-F](https://doi.org/10.1016/0927-7757(93)80240-F).
94. Fredrick E, Walstra P, Dewettinck K: **Factors governing partial coalescence in oil-in-water emulsions.** *Adv Colloid Interface Sci* 2010, **153**:30–42, <https://doi.org/10.1016/j.cis.2009.10.003>.
95. Arima S, Ueji T, Ueno S, Ogawa A, Sato K: **Retardation of crystallization-induced destabilization of PMF-in-water emulsions with emulsifier additives.** *Colloids Surf, B* 2007, **55**:98–106, <https://doi.org/10.1016/j.colsurfb.2006.11.025>.
96. Moens K, Tzompa-Sosa DA, Van de Walle D, Van der Meeren P, Dewettinck K: **Influence of cooling rate on partial coalescence in natural dairy cream.** *Food Res Int* 2019, **120**:819–828, <https://doi.org/10.1016/j.foodres.2018.11.044>.
97. Degner BM, Olson KM, Rose D, Schlegel V, Hutkins R, McClements DJ: **Influence of freezing rate variation on the microstructure and physicochemical properties of food emulsions.** *J Food Eng* 2013, **119**:244–253, <https://doi.org/10.1016/j.jfoodeng.2013.05.034>.
98. Moens K, Masum AKM, Dewettinck K: **Tempering of dairy emulsions: partial coalescence and whipping properties.** *Int Dairy J* 2016, **56**:92–100, <https://doi.org/10.1016/j.idairyj.2016.01.007>.
99. Fredrick E, Heyman B, Moens K, Fischer S, Verwijlen T, Moldenaers P, Van der Meeren P, Dewettinck K: **Monoacylglycerols in dairy recombined cream: II. The effect on partial coalescence and whipping properties.** *Food Res Int* 2013, **51**:936–945, <https://doi.org/10.1016/j.foodres.2013.02.006>.
100. Denkov N, Tcholakova S, Lesov I, Cholakova D, Smoukov SK: **Self-shaping of oil droplets via the formation of intermediate rotator phases upon cooling.** *Nature* 2015, **528**:392–395, <https://doi.org/10.1038/nature16189>.  
The “drop self-shaping” phenomenon is described in detail and is related to the presence of intermediate rotator phases in cooled alkane emulsions. The general drop shape evolutionary scheme is described and the main factors for process control are outlined.
101. Cholakova D, Denkov N, Tcholakova S, Lesov I, Smoukov SK: **Control of drop shape transformations in cooled emulsions.** *Adv Colloid Interface Sci* 2016, **235**:90–107, <https://doi.org/10.1016/j.cis.2016.06.002>.
102. Denkov N, Cholakova D, Tcholakova S, Smoukov SK: **On the mechanism of drop „self-shaping“ phenomenon in cooled emulsions.** *Langmuir* 2016, **32**:7985–7991, <https://doi.org/10.1021/acs.langmuir.6b01626>.
103. Tcholakova S, Valkova Z, Cholakova D, Vinarov Z, Lesov I, Denkov N, Smoukov SK: **Efficient self-emulsification via cooling-heating cycles.** *Nat Commun* 2017, **8**:15012, <https://doi.org/10.1038/ncomms15012>.  
New approach for spontaneous drop fragmentation (self-emulsification) in cooled emulsions is described. Three distinct mechanisms are observed by optical microscopy and, on this basis, the main control factors are discussed.
104. Valkova Z, Cholakova D, Tcholakova S, Denkov N, Smoukov SK: **Mechanisms and control of self-emulsification upon freezing and melting of dispersed alkane drops.** *Langmuir* 2017, **33**:12155–12170, <https://doi.org/10.1021/acs.langmuir.7b02048>.
105. Cholakova D, Denkov N: **Rotator phases in alkane systems: in bulk, surface layers and micro/nano-confinements.** *Adv Colloid Interface Sci* 2019, **269**:7–42, <https://doi.org/10.1016/j.cis.2019.04.001>.  
Comprehensive review of the rotator phase formation in alkane systems, including those formed in surface layers. The review emphasizes that, although often called by different names, the rotator phases are ubiquitous in nature and technology.
106. Cholakova D, Denkov N, Tcholakova S, Valkova Z, Smoukov SK: **Multilayer formation in self-shaping emulsion droplets.** *Langmuir* 2019, **35**:5484–5495, <https://doi.org/10.1021/acs.langmuir.8b02771>.
107. Haas PA, Goldstein RE, Smoukov SK, Cholakova D, Denkov N: **Theory of shape-shifting droplets.** *Phys Rev Lett* 2017, **118**:088001, <https://doi.org/10.1103/PhysRevLett.118.088001>.
108. Haas PA, Cholakova D, Denkov N, Goldstein RE, Smoukov SK: **Shape-shifting polyhedral droplets.** *Phys Rev Res* 2019, **1**:023017, <https://doi.org/10.1103/PhysRevResearch.1.023017>.
109. Guttman S, Sapir Z, Schultz M, Butenko A, Ocko B, Deutsch M, Sloutskin E: **How faceted liquid droplets grow tails.** *Proc Natl Acad Sci U S A* 2016, **113**:493–496, <https://doi.org/10.1073/pnas.1515614113>.  
This is the first paper in the recent series of studies on the drop-shaping phenomena by Guttman et al. in which the monolayer freezing is proposed as the main process leading to drop faceting and to the other drop-shape transformations.
110. Guttman S, Sapir Z, Ocko B, Deutsch M, Sloutskin E: **Temperature-tuned faceting and shape changes in liquid alkane droplets.** *Langmuir* 2017, **33**:1305–1314, <https://doi.org/10.1021/acs.langmuir.6b02926>.

111. Sharker KK, Islam Md N, Das S: **Counterion effect on Krafft temperature and related properties of octadecyltrimethylammonium bromide.** *J Surfact Deterg* 2017, **20**: 923–932, <https://doi.org/10.1007/s11743-017-1957-5>.
112. Lesov I, Valkova Z, Vassileva E, Georgiev GS, Ruseva K, Simeonov M, Tcholakova S, Denkov ND, Smoukov SK: **Bottom-up synthesis of polymeric micro- and nanoparticles with regular anisotropic shapes.** *Macromolecules* 2018, **51**: 7456–7462, <https://doi.org/10.1021/acs.macromol.8b00529>.
113. Marin O, Alesker M, Guttman S, Gershinsky G, Edri E, Shpaisman H, Guerra RE, Zitoun D, Deutsch M, Sloutskin E: **Self-faceting of emulsion droplets as a route to solid icosahedra and other polyhedra.** *J Colloid Interface Sci* 2019, **538**: 541–545, <https://doi.org/10.1016/j.jcis.2018.11.111>.
114. Kovacic F, Okur H, Smolentsev N, Scheu R, Roke S: **Hydration mediated interfacial transitions on mixed hydrophobic/hydrophilic nanodroplet interface.** *J Chem Phys* 2018, **149**: 234704, <https://doi.org/10.1063/1.5035161>.
115. Ralla T, Salminen H, Tuosto J, Weiss J: **Formation and stability of emulsions by Yucca saponin extract.** *Int J Food Sci Technol* 2017, **53**:1381–1388, <https://doi.org/10.1111/ijfs.13715>.
116. Yang Y, Leser ME, Sher AA, McClements DJ: **Formation and stability of emulsions using a natural small molecule surfactant: Quillaja saponin (Q-Naturale®).** *Food Hydrocolloids* 2013, **30**:589–596, <https://doi.org/10.1016/j.foodhyd.2012.08.008>.
117. Bai L, Huan S, Gu J, McClements DJ: **Fabrication of oil-in-water nanoemulsions by dual-channel microfluidization using natural emulsifiers: saponins, phospholipids, proteins, and polysaccharides.** *Food Hydrocolloids* 2016, **61**:703–711, <https://doi.org/10.1016/j.foodhyd.2016.06.035>.
118. Chen X-W, Yang D-X, Zou Y, Yang X-Q: **Stabilization and functionalization of aqueous foams by Quillaja saponin-coated nanodroplets.** *Food Res Int* 2017, **99**:679–687, <https://doi.org/10.1016/j.foodres.2017.06.045>.  
Saponin surfactant is used to create a hierarchical structure of aerated nano-emulsions with high stability and tunable rheological properties. Several potential applications of this type of systems are discussed.
119. Wan Z, Sun Y, Ma L, Zhou F, Guo J, Hu S-Q, Yang X-Q: **Long-lived thermoresponsive emulsion foams stabilized by self-assembled saponin nanofibrils and fibrillar network.** *Langmuir* 2018, **34**:3971–3980, <https://doi.org/10.1021/acs.langmuir.8b00128>.